Simulating the Global Distribution of Nitrogen Isotopes in the Ocean

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Abstract

We present a new nitrogen isotope model incorporated into the three-dimensional ocean component of a global Earth System Climate Model designed for millennial timescale simulations. The model includes prognostic tracers for the two stable nitrogen isotopes, $^{14}$N and $^{15}$N, in the nitrate ($\text{NO}_3^-$), phytoplankton, zooplankton, and detritus variables of the marine ecosystem model. The isotope effects of algal NO$_3^-$ uptake, nitrogen fixation, water column denitrification, and zooplankton excretion are considered as well as the removal of NO$_3^-$ by sedimentary denitrification. A global database of $\delta^{15}$NO$_3^-$ observations is compiled from previous studies and compared to the model results on a regional basis where sufficient observations exist. The model is able to qualitatively and quantitatively reproduce many of the observed patterns such as high subsurface values in water column denitrification zones and the meridional and vertical gradients in the Southern Ocean. The observed pronounced subsurface minimum in the Atlantic is underestimated by the model presumably owing to too little simulated nitrogen fixation there. Sensitivity experiments reveal that algal NO$_3^-$ uptake, nitrogen
fixation and water column denitrification have the strongest effects on the simulated distribution of nitrogen isotopes, whereas the effect from zooplankton excretion is weaker. Both water column and sedimentary denitrification also have important indirect effects on the nitrogen isotope distribution by lowering the fixed nitrogen inventory, which creates an ecological niche for nitrogen fixers and, thus, stimulates additional N$_2$ fixation in the model. Important model deficiencies are identified, and strategies for future improvement and possibilities for model application are outlined.

1. Introduction

Bioavailable nitrogen (fixed-N) is one of the major limiting nutrients for algal photosynthesis, which drives the sequestration of CO$_2$ from the surface ocean and atmosphere into the deep ocean via the sinking of organic matter. Changes in this so-called 'biological pump' have been hypothesized to account for a significant amount of the glacial-interglacial fluctuations in atmospheric CO$_2$ [McElroy, 1982; Falkowski, 1997]. However, the relative contributions of the biological and physical carbon pumps to CO$_2$ variations remain controversial. The size of the oceanic fixed-N inventory, which regulates the strength of the biological pump, is controlled by different biogeochemical processes that are difficult to constrain quantitatively in a global budget [Codispoti, 2007]. Nitrogen isotopes (both in dissolved and organic N species) in the water column and sea floor sediments are sensitive indicators of those processes [Brandes and Devol, 2002; Altabet, 2007].

Many N-transformational processes alter the ratio of the two stable forms of the nitrogen isotopes, $^{14}$N and $^{15}$N, differently, a process referred to as fractionation. Resulting variations in N isotopic composition can be described as deviations in $^{15}$N/$^{14}$N ratio from an accepted standard

$$\delta^{15}N = [(^{15}N/^{14}N) / R_{std} - 1] \times 1000 \quad , \quad (1)$$
where $R_{std}$ is the $^{15}\text{N}/^{14}\text{N}$ ratio of atmospheric $\text{N}_2$ gas. Isotope fractionation can occur due to kinetic processes (i.e., different reaction rates for isotopes in a reactant-product stream). It generally results in the enrichment of the heavier $^{15}\text{N}$ isotope in the reaction substrate, and its depletion in the product. For example, preferential discrimination against $^{15}\text{N}$ relative to $^{14}\text{N}$ during algal $\text{NO}_3^-$ assimilation results in net enrichment of $^{15}\text{N}$ in the residual $\text{NO}_3^-$ and net depletion of $^{15}\text{N}$ in organic matter (OM). The degree of isotopic discrimination, or fractionation, for each process can be quantified with an enrichment factor, $\varepsilon = (^{14}\text{k} / ^{15}\text{k} - 1) \cdot 1000$, where $k$ is the specific reaction rate for each isotope [Mariotti et al., 1981].

The predominant source and sink terms of the oceanic fixed-N inventory, $\text{N}_2$ fixation and denitrification, respectively, have their own distinct effects on the marine N isotope budgets. $\text{N}_2$-fixing bacteria (diazotrophs) introduce bioavailable N into the ocean close to that of atmospheric $\text{N}_2$ ($\delta^{15}\text{N} \approx −2 − 0\%$) [Delwiche and Steyn, 1970; Minagawa and Wada, 1986; Macko et al., 1987; Carpenter et al., 1997; Montoya et al., 2002]. Denitrification occurs under suboxic conditions ($\text{O}_2 < 5 \mu\text{mol/kg}$) in the water column and in the sea floor sediments. Here, microbes use $\text{NO}_3^-$ instead of $\text{O}_2$ as the electron acceptor during respiration and convert it to gaseous forms of N ($\text{N}_2\text{O}$ and $\text{N}_2$), which can then escape to the atmosphere [Codispoti and Richards, 1976]. The volume and distribution of suboxic water is affected by the temperature-dependent solubility of $\text{O}_2$ at the surface and the rate of subduction of oxygen-saturated water masses to greater depths, as well as the amount of organic matter that remineralizes in the ocean interior, both of which are sensitive to changes in climate.

Denitrifiers preferentially consume $^{14}\text{NO}_3^-$ leaving the residual oceanic $\text{NO}_3^-$ pool strongly enriched in the heavier $^{15}\text{N}$, with N isotope enrichment factors between 20 – 30\% [Cline and Kaplan, 1975; Liu...
Sedimentary denitrification is generally limited by the amount of NO$_3^-$ that diffuses into the reactive zones within the sediments. Therefore, it consumes nearly all of the influxing NO$_3^-$ available, leaving nearly unaltered $\delta^{15}$N values in the overlying waters [Brandes and Devol, 1997, 2002; Sigman et al., 2003; Lehmann et al., 2004, 2007]. The average oceanic $\delta^{15}$NO$_3^-$ value near 5‰ [Sigman et al., 1997, 1999] can be interpreted as the balance between the isotope effects of water column denitrification, sedimentary denitrification, and N$_2$ fixation [Brandes and Devol, 2002; Deutsch et al., 2004; Galbraith et al., 2004; Altabet, 2007].

The $\delta^{15}$N signal in the water column and sea floor sediments is also affected by fractionation processes within the food chain. Marine algae preferentially assimilate the lighter $^{14}$N into their biomass with a range of enrichment factors estimated in the field between 4 – 15‰ [Wada, 1980; Altabet et al., 1991; Altabet and Francois, 1994; Altabet et al., 1999; Sigman et al., 1999; Altabet and Francois, 2001; Karsh et al., 2003; DiFiore et al., 2006]. Nitrogen is not lost or gained from the ocean during algal NO$_3^-$ assimilation, but the spatial separation between net assimilation and remineralization can cause a trend of decreasing $\delta^{15}$NO$_3^-$ with depth. Distinguishing between the different isotope effects remains a challenge, especially in regions where multiple N-transformational processes are occurring within close proximity.

This study adds a dynamic nitrogen isotope module to an existing global ocean-atmosphere-sea ice-biogeochemical model designed for millennial timescale simulations. We provide a detailed description of the nitrogen isotope model and an assessment of its skill in reproducing present day $\delta^{15}$NO$_3^-$ observations. Comparison of model results with $\delta^{15}$N observations will also be used to help to quantify processes that affect the global oceanic distribution of $\delta^{15}$N. Sensitivity experiments illustrate
the individual isotope effects of different processes on the spatial distribution of the N isotopes. In combination with measurements in ocean sediments and in the water column, the model can be a tool to better understand variations of $\delta^{15}$N and the nitrogen cycle in the past and present.

2. Model Description

2.1 Physical Model

The physical model is based on the University of Victoria Earth System Climate Model [Weaver et al., 2001], version 2.8. It includes a global, three-dimensional general circulation model of the ocean (Modular Ocean Model 2) with physical parameterizations such as diffusive mixing along and across isopycnals, eddy induced tracer advection [Gent and McWilliams, 1990] and a scheme for the computation of tidally induced diapycnal mixing over rough topography [Simmons et al., 2004]. Nineteen vertical levels are used with a horizontal resolution of 1.8°x3.6°. To improve the simulation of equatorial currents, we have increased the meridional resolution in the tropics to 0.9° (between 10°S and 10°N and smoothly transitioning to 1.8° at 20°N/S) and added an anisotropic viscosity scheme [Large et al., 2001]. A more detailed description of this parameterization and its effect on the equatorial circulation is provided in Supplementary Material 1. To account for the overestimated ventilation in the North Pacific, an artificial stratifying force equal to 0.05 Sv of freshwater is applied over the surface north of 55° in the Pacific and compensated elsewhere. A two dimensional, single level energy-moisture balance model of the atmosphere and a state-of-the-art dynamic-thermodynamic sea ice model are used, forced with prescribed NCEP/NCAR monthly climatological winds.

2.2 Marine Ecosystem Model

The marine ecosystem model is an improved version of the NPZD (Nutrient, Phytoplankton, Zooplankton, Detritus) ecosystem model of Schmittner et al. [2008] (Figure 1). The organic variables
include two classes of phytoplankton, N\textsubscript{2}-fixing diazotrophs (P\textsubscript{D}) and a NO\textsubscript{3}\textsuperscript{-} assimilating algal phytoplankton class (P\textsubscript{A}), as well as zooplankton (Z) and organic detritus (D). The inorganic variables include dissolved oxygen (O\textsubscript{2}) and two nutrients, nitrate (NO\textsubscript{3}\textsuperscript{-}) and phosphate (PO\textsubscript{4}\textsuperscript{3-}), both of which are consumed by phytoplankton and remineralized in fixed elemental ratios (R\textsubscript{N:P} = 16, R\textsubscript{O:P} = 170).

We note, though, that diazotrophs have been found to have R\textsubscript{N:P} as high as 50:1 (e.g., Letelier and Karl, [1996, 1998]). This simplification is one of the reasons why the nitrogen surplus N' = NO\textsubscript{3}\textsuperscript{-}−16PO\textsubscript{4}\textsuperscript{3-} is generally underestimated in surface waters in the model (Figure S2-1). In addition to water column denitrification and N\textsubscript{2} fixation, we now include a parameterization for sedimentary denitrification (see Supplementary Material 2, equation S2-11 and Figure 2), based on the flux of organic carbon into the sea floor sediments [Middleburg et al., 1996]. Since the model underestimates coastal upwelling, which drives large fluxes of organic carbon to the sea floor sediments, this parameterization is tuned to fit the global mean \(\delta^{15}\text{NO}_3^-\) observations. The complete marine ecosystem model description is provided in Supplementary Material 2. A comparison of the global distribution of NO\textsubscript{3}\textsuperscript{-}, O\textsubscript{2}, and N' with World Ocean Atlas 2005 (WOA05) observations is shown in Figure S2-1.

Water column denitrification is present in three main locations of the present-day oceans: the Eastern Tropical North Pacific (ETNP), the Eastern Tropical South Pacific (ETSP) and the Arabian Sea. Deficiencies in the physical circulation model simulate suboxic water in only one of these locations, the ETNP (Figure S2-1b). The physical circulation model integrates coastal upwelling over a horizontal extent that is too large (due to its coarse resolution), which results in the underestimation of upwelling, export production, and the remineralization of organic matter at depth. This bias leads to too high O\textsubscript{2} concentrations, larger than required for water column denitrification, in the ETSP and the Arabian Sea. Suboxia in the so-called “shadow zone” of the ETNP is simulated better and investigated more in Section 4.2. In the model, some water column denitrification also occurs in the Bay of Bengal.
and off SW Africa (Figure 2C), which has not been observed in the real ocean. However, the anammox reaction, which also eliminates fixed-N in the water column, has been found to occur off SW Africa \cite{Kuypers:2005, Kuypers:2006}. Navqi et al. [1996] measured low decomposition rates in the Bay of Bengal. Effective ballasting and scavenging of organic matter by the massive riverine input of terrestrial matter, an effect not included in the model, may prevent water column denitrification in the Bay of Bengal, which is close to suboxic.

Diazotroph’s grow according to the same principles as algal phytoplankton in the model (see Supplementary Material 2), but we also account for some of their different characteristics. \(\text{N}_2\) fixation breaks down of the triple N bond of \(\text{N}_2\), which is energetically more costly than assimilating fixed-N. Therefore, in the model, the growth rate of diazotrophs is lower than that of algal phytoplankton. It is zero in waters cooler than 15\(^\circ\)C and increases 50\% slower with temperature than the growth rate of algal phytoplankton. Diazotrophs are not limited by \(\text{NO}_3^-\) and will thrive in waters that are N-deficient (i.e., low N\(^’\) as a result of denitrification) in which sufficient P and Fe are available. Denitrification and the propagation of N-deficient waters into the shallow thermocline by physical transport processes creates an ecological niche for diazotrophs in the model, which stimulates \(\text{N}_2\) fixation [Tyrell, 1999].

One of the most important and best studied diazotrophs, \textit{Trichodesmium}, also has large iron (Fe) requirements for growth [Sanudo-Willhelmy et al., 2001]. Diazotrophs may depend on aeolian Fe in oligotrophic waters because deep pycnocline inhibits upward mixing of subsurface Fe-replete waters into the euphotic zone [Letelier and Karl, 1996; Falkowski, 1997]. Therefore, their growth rate is reduced by an additional 50\% where estimated rates of aeolian dissolved Fe deposition are very low (< \(\sim 10 \text{ mmol m}^{-2} \text{ yr}^{-1}\)) (Figure 2A), primarily throughout the Tropical South Pacific [Fan et al., 2006]. This is a simple parameterization of Fe limitation of diazotrophy and its full effects are described
elsewhere [Somes, 2009; Somes and Schmittner, in prep.]. The majority of N₂ fixation in the model occurs in oligotrophic waters 'downstream' of denitrification zones where sufficient Fe exists (i.e., via aeolian Fe deposition) (Figure 2B). The pattern of N₂ fixation—such as high values in the tropical/subtropical North Pacific, the western tropical/subtropical South Pacific, the western tropical/subtropical South Atlantic, the tropical/subtropical North Atlantic and the Indian Ocean—is mostly consistent with direct observations (e.g. Karl et al., [2002]), with estimates based on the observed NO₃⁻ deficit and simulated circulation [Deutsch et al., 2007], as well as with results from a more complex ecosystem model [Moore et al., 2007]. However, N₂ fixation in our model does not extend northward of 20–25°N in the North Pacific and North Atlantic, whereas some observations show N₂ fixation as far north as 35–40°N.

2.3 Nitrogen Isotope Model

The nitrogen isotope model simulates the distribution of the two stable nitrogen isotopes, ¹⁴N and ¹⁵N, in all N species throughout the global ocean that are included in the marine ecosystem model. Five prognostic variables of δ¹⁵N are embedded within the marine ecosystem model for all species containing nitrogen: NO₃⁻, diazotrophs, algal phytoplankton, zooplankton and organic detritus (Figure 1). The 'isotope effect' is referred to in the following as the effect that each process has on the respective oceanic isotopic N pool, which depends on the δ¹⁵N value of the substrate, the process-specific enrichment factor (ε), and the degree of utilization (u_substrate) of the substrate during the reaction:

\[ \delta^{15}N_{\text{product}} = \delta^{15}N_{\text{substrate}} - \varepsilon (1 - u_{\text{substrate}}) \]  

where \( u_{\text{substrate}} \) is the fraction of the initial substrate used in the reaction. For example, if all of the
available substrate is consumed in the reaction (i.e., $u_{\text{substrate}} = I$), the product will incorporate the $\delta^{15}\text{N}$ value of the substrate, nullifying any potential fractionation. However, if the rate of utilization is low (i.e., $u_{\text{substrate}} \sim 0$), the product will incorporate a relatively light $\delta^{15}\text{N}$ value compared to the substrate by the designated enrichment factor (Table 1).

The processes in the model that fractionate nitrogen isotopes are algal NO$_3^-$ assimilation ($\varepsilon_{\text{ASSIM}} = 5\%$), zooplankton excretion ($\varepsilon_{\text{EXCR}} = 6\%$), and water column denitrification ($\varepsilon_{\text{WCD}} = 25\%$) (Table 1). Fractionation results in the isotopic enrichment of the more reactive, thermodynamically preferred, light $^{14}\text{N}$ into the product of each reaction by a process-specific fractionation factor. For a detailed discussion of nitrogen isotope fractionation dynamics see Mariotti et al. [1981]. Although little fractionation occurs during N$_2$ fixation in the model, it has an important effect on $\delta^{15}\text{N}$ by introducing relatively light atmospheric N$_2$ ($\delta^{15}\text{N} = 0\%$) into the oceanic fixed-N inventory. Sedimentary denitrification also has been observed to have little effect on the oceanic isotopic N pool because denitrifiers consume nearly all NO$_3^-$ diffusing into the sediments [Brandes and Devol, 1997, 2002; Lehmann et al., 2004, 2007]. In the model, there is no fractionation during sedimentary denitrification ($\varepsilon_{\text{SD}} = 0\%$), although this is a simplification of observations [Lehmann et al., 2007]. Fractionation during the remineralization of organic matter is not included in the model. The complete nitrogen isotope model description is provided in Appendix A.

3. Nitrogen Isotope Model Results

The model simulates complex spatial patterns of $\delta^{15}\text{NO}_3^-$ and $\delta^{15}\text{N}$-organic matter (OM) throughout the global ocean (top panels of Figure 3). Patterns of surface $\delta^{15}\text{NO}_3^-$ and subsurface $\delta^{15}\text{N}$-OM are very similar but values are offset by two processes. First, as much as 5% offset due to fractionation during NO$_3^-$ uptake by phytoplankton and second, by fractionation during zooplankton excretion, which
increases the $\delta^{15}N$-OM through zooplankton mortality (Figure 1). High $\delta^{15}N$ values (>15‰) are simulated in the eastern subtropical gyres, where surface NO$_3^-$ is depleted, and in regions in close proximity to simulated suboxic zones in the Eastern Pacific, Bay of Bengal, and Eastern Atlantic (again, note that water column denitrification is has not been observed in the Bay of Bengal and Eastern Atlantic). A clear inter-hemispheric asymmetry appears between the subtropical gyres of the Pacific and Atlantic with higher values of 14-20‰ simulated in the southern hemisphere and smaller values of 10-14‰ in the northern hemisphere. More intermediate $\delta^{15}N$ values of 4 - 8‰ are found at high latitudes and near the equator where nutrient utilization is incomplete. $\delta^{15}N$ minima (<4‰) are located in the western tropical/subtropical ocean basins, where N$_2$ fixation occurs in the model (Figure 2B). The remainder of this section presents a more quantitative description of the contributions of individual processes to these relatively complex spatial patterns of $\delta^{15}NO_3^-$ and $\delta^{15}N$-OM.

Figure 3 illustrates results from the full model (CTRL) that includes all isotope effects (top panels) together with results from sensitivity experiments designed to isolate the effects of individual processes (bottom panels) on the global $\delta^{15}N$ distribution. This is accomplished by removing the isotope effect of one process per experiment and then calculating the difference ($\Delta \delta^{15}N$) with CTRL. In the “NO$_3^-$ Assimilation” and “Excretion” experiments, the enrichment factors $\varepsilon_{\text{ASSIM}}$ and $\varepsilon_{\text{EXCR}}$, respectively, are set to zero. In the “N$_2$ fixation” experiment the diazotroph’s N isotope ratio is set equal to that of other phytoplankton at each location. In the “Water Column Denitrification” and “Sedimentary Denitrification” experiments, the entire process is switched off (thereby changing the global N inventory). These latter experiments also show the indirect effect that both denitrification processes have on $\delta^{15}N$ through the stimulation of N$_2$ fixation. In all other isotope effect experiments, the total N inventory does not change.
3.1 Algal NO$_3^-$ Assimilation

As phytoplankton preferentially assimilate $^{14}$NO$_3^-$ into organic matter, they leave the residual inorganic N pool enriched in $^{15}$NO$_3^-$, which creates an offset up to 5‰ between surface $\delta^{15}$NO$_3^-$ and surface $\delta^{15}$N-OM that sinks towards the sea floor (“NO$_3^-$ Assimilation” experiment, Figure 3). The surface NO$_3^-$ utilization effect depends on the extent to which NO$_3^-$ is consumed with respect to its physical supply. When NO$_3^-$ utilization is low, which occurs in High Nitrate Low Chlorophyll (HNLC) regions such as the Southern Ocean, the northern North Pacific, and the eastern equatorial Pacific, surface $\delta^{15}$NO$_3^-$ is controlled mainly by the source of $\delta^{15}$NO$_3^-$ being supplied to the surface. Algae will fractionate NO$_3^-$ during assimilation near their given enrichment factor in these N-replete waters so that the expected 5‰ difference between $\delta^{15}$NO$_3^-$ and $\delta^{15}$N-OM is almost fully expressed (i.e., $\delta^{15}P_A = \delta^{15}$NO$_3^-$ - $\epsilon_{\text{ASSIM}}$ with $u_{\text{ASSIM}} = 0$ in Equation 2). Thus, NO$_3^-$ utilization in HNLC regions has a small effect on surface $\delta^{15}$NO$_3^-$ but has a much larger effect on $\delta^{15}$N-OM. This is perhaps most obvious in the Southern Ocean and in the northern North Pacific where $\Delta \delta^{15}$NO$_3^-$ is small, whereas $\Delta \delta^{15}$N-OM is strongly negative.

A different response is observed in oligotrophic regions where surface NO$_3^-$ is depleted. Once the algae consume nearly all available NO$_3^-$ (which itself becomes enriched in $^{15}$N), they acquire the same N isotope signature from the source NO$_3^-$ (i.e., $\delta^{15}P_A = \delta^{15}$NO$_3^-$ with $u_{\text{ASSIM}}$ approaching 1). This drives the high $\delta^{15}$N values in both NO$_3^-$ and OM in the subtropics with maxima in the eastern poleward edges of the gyres (Figure 3). Although $\delta^{15}$NO$_3^-$ values are very high there, they have a small effect on $\delta^{15}$N elsewhere because NO$_3^-$ concentrations are very low. For instance, when low NO$_3^-$ water mixes with nearby water with significantly higher NO$_3^-$, the resulting $\delta^{15}$NO$_3^-$ value will be weighted towards the water parcel containing more NO$_3^-$ (see also Deutsch et al., [2004]). This ‘dilution effect’ prevents high $\delta^{15}$NO$_3^-$ values in regions with high surface NO$_3^-$ utilization from having
a large impact on the $\delta^{15}$NO$_3^-$ signature across the nitrcline.

3.2 Denitrification

Water column denitrification only occurs at depth but its isotope effect can reach the surface due to upwelling and vertical mixing of water enriched in $^{15}$NO$_3^-$. It has a large enrichment factor and displays a very strong N isotope effect in close proximity to the simulated suboxic zones in the Eastern Pacific, Bay of Bengal, and Eastern Atlantic (“Water Column Denitrification” experiment in Figure 3). The unresolved poleward undercurrents along the western continental margin of the Americas—which could, in the real world, propagate high $\delta^{15}$NO$_3^-$ away from the subsurface suboxic zones [Kineast et al., 2002]—may restrict the simulated water column denitrification isotope effect too much to regions proximal to the suboxic zones. Both water column and sedimentary denitrification also indirectly lead to lower $\delta^{15}$NO$_3^-$ values “downstream” of denitrification zones by creating more N-deficient water, which stimulates additional N$_2$ fixation. This negative feedback also decreases the horizontal extension of high $\delta^{15}$NO$_3^-$ signature originating from suboxic zones, because N$_2$ fixation introduces much lower $\delta^{15}$N into the ocean.

3.3 N$_2$ Fixation

The addition of newly fixed, isotopically light atmospheric N$_2$ ($\delta^{15}$N$_2 = 0$) by diazotrophs is the reason for the low $\delta^{15}$N values in the western tropical/subtropical ocean basins. Since denitrification is the only process in the model that creates N-deficient water, and therefore an ecological niche for diazotrophs, the majority of N$_2$ fixation in the model occurs “downstream” of denitrification zones after phytoplankton have consumed all remaining surface NO$_3^-$ and where sufficient aeolian Fe deposition exists. This low $\delta^{15}$NO$_3^-$ signature is evident in the subtropical North/South Pacific, the subtropical North/South Atlantic, and the Bay of Bengal (“N$_2$ Fixation” experiment, Figure 3).
According to our model results, the N isotope effect of excretion has a smaller influence on the simulated distribution of $\delta^{15}N$ in the global ocean (“Excretion” experiment, Figure 3) compared to the other processes discussed above. Its strongest effect is observed in the subtropical South Pacific, where $NO_3^-$ is very low and excretion significantly contributes to the $NO_3^-$ pool by introducing relatively low $\delta^{15}NO_3^-$. Low latitude surface waters elsewhere are generally about 1-4‰ lighter due to fractionation during excretion, with little spatial gradients. At high latitudes the effect on $\delta^{15}NO_3^-$ is very small. We note that this N isotope effect is sensitive to the parameterization for excretion used in this marine ecosystem model version. The excretion rate was tuned so that $\delta^{15}N$-zooplankton is enriched by $\sim$3.4‰ relative to phytoplankton [Minagawa and Wada, 1984].

4. Model Evaluation

The relatively small number of $\delta^{15}N$ observations and the sparse spatial and temporal coverage make a full global model assessment difficult. However, certain regions have been sampled sufficiently to provide a meaningful comparison with the model results. All observations presented here are interpolated horizontally onto a 0.9° x 1.8° grid using a Gaussian weighted algorithm. The 33 depth levels are consistent with WOA05 and a linear interpolation is used for depths of missing data if nearby data exist. A global database of $\delta^{15}NO_3^-$ measurements has thus been constructed and is available for download (http://mgg.coas.oregonstate.edu/~andreas/Nitrogen/n15database.html). Figure 4 shows the annually averaged global distribution of measured $\delta^{15}NO_3^-$, averaged over 200 – 300 m depth to illustrate the spatial coverage. Seasonal sampling biases exist depending on the region. More details on the data sets can be found in the respective ocean-region subsections that follow. Comparisons are presented for the Southern Ocean (Indian-Pacific sector), the Eastern Tropical North Pacific, the
Central Equatorial Pacific and the Subtropical North Atlantic. Other regions with available $\delta^{15}\text{NO}_3^-$ observations included in the dataset but not discussed in the text are the Bering Sea [Lehmann et al., 2005], the Northeast Pacific [Galbraith, 2006], the Arabian Sea [Altabet et al., 1999] and the eastern Pacific sector of the Southern Ocean [Sigman et al., 1999].

Global measures of model performance for $\delta^{15}\text{NO}_3^-$ are presented in Table 2. These measures should be interpreted taken into account the highly localized nature of some of the processes as well as the limited regions covered by the database. A displacement in the location of denitrification, for example, will lead to a large decrease in the correlation coefficient and a large increase in the RMS errors. The CTRL model has a correlation coefficient of 0.68, implying that the model explains 46% of the variance in the observations. The decrease of the correlation coefficient and the increase of the RMS error due to the neglection of a particular process can be regarded as the importance that this process plays in explaining the global $\delta^{15}\text{NO}_3^-$ observations of the database. The correlation coefficient measures the pattern of variability and neglects the absolute values, whereas the RMS error considers the deviation of the model from the observations in absolute values. Neglecting water column denitrification leads to the largest decrease in the correlation coefficient to 0.29 and to the second largest increase in the RMS error after N$_2$ fixation. Neglecting algal NO$_3^-$ assimilation also leads to a large decrease in the correlation coefficient and to a large increase in the RMS error. If N$_2$ fixation or sedimentary denitrification is not included, then the correlation coefficients decrease similarly, but exclusion of N$_2$ fixation causes a larger increase in the RMS error relative to sedimentary denitrification. Excretion has the weakest effect on the observed $\delta^{15}\text{NO}_3^-$ distribution in the model. Then, according to these measures, water column denitrification is the most important process determining the global $\delta^{15}\text{NO}_3^-$ distribution of the database, followed by algal NO$_3^-$ assimilation. N$_2$ fixation is more important than sedimentary denitrification, whereas excretion is the least important.
The Southern Ocean represents a critical region of biogeochemical cycling in the ocean because it is the largest open ocean region with incomplete drawdown of the major nutrients. This results in an excess amount of CO$_2$ at the surface, which is released to the atmosphere (under pre-industrial conditions). The degree to which surface nutrients are utilized here may have profound impacts on ocean-atmosphere exchanges of CO$_2$. Figure 5 shows a comparison with observations recorded in the region [Sigman et al., 1999; Altabet and Francois, 2001; DiFiore et al., 2006]. This data sub-set compiles observations from 8 cruises covering various seasons. Since all cruises do not cover the same location, some seasonal biases can be expected, yet, we still decided to use annual averages for maximum spatial coverage. The model does not simulate interannual variability due to the prescribed monthly climatological winds and temporally constant biogeochemical parameters.

Qualitatively, the inverse trend of increasing $\delta^{15}$NO$_3^-$ with decreasing NO$_3^-$ (Figure 5a) is reproduced by the model. However, the slope is underestimated suggesting that the enrichment factor for algal NO$_3^-$ assimilation used in the model ($\varepsilon_{\text{ASSIM}} = 5\%$) is too low, in agreement with DiFiore et al. [2006] that suggests at least 7‰. The simulated vertical gradient is in good agreement with the observations. Deep water $\delta^{15}$NO$_3^-$ at 2000 m depth is around 5‰ and slowly increasing throughout the lower pycnocline to around 6‰ at 500 m depth. The model slightly overestimates $\delta^{15}$NO$_3^-$ between 200 m and 400 m depth, whereas near surface values are slightly underestimated.

A large discrepancy between simulated and observational $\delta^{15}$NO$_3^-$ is apparent in surface waters north of 40°S off the southern coast of Australia (Figure 5c). This bias is due to the fact that the model overestimates the utilization of surface NO$_3^-$ relative to observations there (Figure 5c). Where the
simulated NO$_3^-$ is almost completely consumed (i.e., NO$_3^-$ < 1 μM) (see Figure 5c-contour line), the
remaining δ$^{15}$NO$_3^-$ values become as high as 18‰. Since none of the existing δ$^{15}$NO$_3^-$ observations
was collected in such low NO$_3^-$ concentrations (Figure 5a), it impossible, at this time, to falsify this
aspect of the N isotope model response. We note this heavy δ$^{15}$NO$_3^-$ signature in these low NO$_3^-$
waters has little effect on δ$^{15}$NO$_3^-$ across the nitracline in the model because the δ$^{15}$N signature of very
low NO$_3^-$ water becomes diluted out as it mixes with much higher NO$_3^-$ water (see Section 3.1).

4.2 Eastern Tropical North Pacific

The ETNP contains the largest suboxic zone in the ocean, where water column denitrification occurs.
The relatively small spatial scale of suboxic zones makes them difficult for the model to simulate
accurately and deficiencies in the coarse resolution physical model prevent it from fully resolving some
important physical processes, especially in coastal regions. Underestimating coastal upwelling (due to
coarse resolution) results in corresponding underestimation of primary production, organic matter
remineralization, and O$_2$ consumption at depth. This is a major reason for overestimated dissolved O$_2$
at depth in areas with significant coastal upwelling (e.g., off Peru and NW Mexico) (Figure S2-1), too
large for water column denitrification to occur. Preliminary experiments suggest that increased vertical
resolution can improve the simulation of productivity and suboxia in the Eastern Tropical South Pacific
(not shown).

The ability to reproduce the equatorial undercurrents that transport relatively oxygen-rich water from
the western basin is also important for the simulation of the Eastern Pacific suboxic zones. The
anisotropic viscosity scheme [Large et al., 2001] improves equatorial dynamics considerably
(Supplementary Material 1). The Pacific Equatorial Undercurrent increases from 0.15 m/s to nearly 0.8
m/s, just slightly weaker than observations, which show velocities near 1 m/s (Figure S1-2). The North
Equatorial Countercurrent in the model also displays lower current velocities than observed, and does not deliver enough oxygen-rich water directly to the ETNP suboxic zone. This is likely the main reason why the simulated suboxic zone is much larger than observed towards the equator (Figures 6c).

Figure 6 shows model $\delta^{15}$NO$_3^-$ compared to observational $\delta^{15}$NO$_3^-$ data collected during November 1999 [Sigman et al., 2005] and October 2000 [Altabet, previously unpublished]. The model captures the general observed trend of increasing $\delta^{15}$NO$_3^-$ as NO$_3^-$ is consumed during water column denitrification (Figure 6a). The model’s too low N:P ratio for diazotrophs may partly explain its incapacity to simulate some of the relatively high N' values of observations. The range of simulated values is also likely to be more limited compared to the observations due to the missing interannual and synoptic climate variability in the model.

Due to the described shortcomings in the physical circulation, the simulated suboxic zone is too large and located too far south (by ~5°) relative to observations (Figure 6c-contour line). This results in higher rates of water column denitrification and higher $\delta^{15}$NO$_3^-$ values, as well as more N-deficient water in the suboxic zone compared to observations. Figure 6b compares the horizontally averaged $\delta^{15}$NO$_3^-$ depth profiles only within the hypoxic zone (O$_2$ < 10 μM) at 300 m (contoured on Figure 6c) to account for the displaced OMZ. Within this region, the model is able to capture the general vertical distribution of $\delta^{15}$NO$_3^-$ seen in the measured data, such as the surface minimum and subsurface maximum.

$\delta^{15}$NO$_3^-$ in the ETNP decreases towards the surface [Cline and Kaplan, 1975; Brandes et al., 1998; Voss et al., 2001; Sigman et al., 2005] suggesting a source of isotopically light N at the surface. Brandes et al. [1998] proposed that in the Arabian Sea as much as 30% of primary production must be
supported by N₂ fixation in order to account for the low surface δ¹⁵NO₃⁻. Other observations also suggest that the decrease in δ¹⁵NO₃⁻ towards the surface is likely due to the fixation of atmospheric N₂ and the subsequent, closely coupled remineralization-nitrification cycle [Sigman et al., 2005]. We test this hypothesis by comparing the observations with the model experiment in which the isotope effect of N₂ fixation is neglected (“No NFIX”). In this case, the model overestimates surface δ¹⁵NO₃⁻ by ~10‰ (Figure 6b) and the surface minimum is not simulated. This experiment demonstrates that the input of isotopically light fixed-N from N₂ fixation in the model best explains the decreasing trend of δ¹⁵NO₃⁻ observations towards the surface. In the model, 20% of the fixed-N loss via denitrification is re-introduced into the surface by N₂ fixation occurring directly above the denitrification zone in the ETNP. The fact that the difference between the subsurface maximum and the near surface minimum is underestimated in the model (4‰ versus 8‰ in the observations) suggests that in the real world the locally re-introduced fraction could be larger than 20%.

4.4 North Atlantic

Uncertainties regarding processes that can affect the nitrogen isotope signal make it challenging to interpret and simulate nitrogen isotopes in the North Atlantic. Estimates of atmospheric N deposition [Duce et al., 2008] and the assimilation-remineralization-nitrification cycle are not well constrained. Although atmospheric N deposition may be significant in this region [Michaels et al., 1996; Lipschultz et al., 2002; Hansell et al., 2004, 2007; Knapp et al., 2005, 2008], its isotopic composition is not well known and therefore is not included in the model at this time. Figure 7 shows the comparison of annual model δ¹⁵NO₃⁻ with available observations from cruises in May 2001, 2004 [Altabet and Montoya, previously unpublished], October 2002 [Knapp et al., 2008], and May 2005 [Bourbounnais et al., 2009]. The model overestimates the δ¹⁵NO₃⁻ values everywhere, by 0.9‰ on average and by 2‰ at 200 m depth, presumably due mostly to the underestimation of N₂ fixation, but possibly also because atmospheric N deposition and/or fractionation during the remineralization of organic matter
are not included. Both of these processes would act to decrease subsurface values of $\delta^{15}$NO$_3^-$.

Underestimated N' in the North Atlantic (Figure S2-1) also indicates too little N$_2$ fixation, but we again note the too low N:P ratio for diazotrophs also contributes to this N' underestimation to some degree.

N$_2$ fixation is most likely underestimated in the model because it does not consider dynamic elemental cycling of the microbial loop. It has been suggested that DOP is more labile relative to DON and recycles through the microbial loop more efficiently, which can help relieve diazotrophs of P limitation in this region [Wu et al., 2000] and enhance N$_2$ fixation. The model is able to reproduce the pattern of low $\delta^{15}$NO$_3^-$ in the thermocline qualitatively, just not quantitatively to the extent present in the observations. Sedimentary denitrification in the North Atlantic stimulates enough N$_2$ fixation in the model to generate a subsurface $\delta^{15}$NO$_3^-$ minimum. When sedimentary denitrification is switched off (“No SD”), the thermocline minimum is not simulated. This suggests that sedimentary denitrification is an important factor influencing N$_2$ fixation in the Subtropical North Atlantic, but not the only factor.

5. Discussion and Conclusions

A new model of nitrogen isotopes has been implemented into the three-dimensional ocean component of a global Earth System Climate Model capable of millennial timescale simulations. Despite some model deficiencies, we have shown that this model can successfully reproduce the general spatial patterns of $\delta^{15}$NO$_3^-$ measured in the ocean. Sensitivity experiments allowed us to isolate the individual N isotope effects of various N-transformational processes on the global distribution of $\delta^{15}$N. Algal NO$_3^-$ assimilation, water column denitrification, and N$_2$ fixation all have strong influences in setting the global patterns of $\delta^{15}$NO$_3^-$ in the ocean, whereas the effect of zooplankton excretion is weaker.

These simulations show that the isotope effect of algal NO$_3^-$ assimilation can drive very large spatial
gradients in both $\delta^{15}$NO$_3^-$ and $\delta^{15}$N-OM depending on the ocean environment (Figure 3). In HNLC areas where surface NO$_3^-$ utilization is low and algae are able to fractionate NO$_3^-$ at their designated enrichment factor, the $\delta^{15}$N-OM signature decreases. However, when NO$_3^-$ utilization is high, the $\delta^{15}$N-OM signature is more similar to the $\delta^{15}$NO$_3^-$ value it consumes because the effective degree of fractionation becomes much lower (see Section 3.1). Surface NO$_3^-$ utilization gradients can transition rapidly, for example due to changes in ocean circulation, and can possibly drive large and rapid changes in $\delta^{15}$NO$_3^-$ and $\delta^{15}$N-OM. The important influence of surface NO$_3^-$ utilization on the global distribution of N isotopes in the model suggests that changes in surface NO$_3^-$ utilization patterns throughout Earth’s history could contribute to large fluctuations in $\delta^{15}$N observed in sediment records, especially near fronts where large surface NO$_3^-$ gradients exist (See also Altabet and Francois [1994]; Farrell et al. [1995]; Francois et al. [1997]; Sigman et al. [1999]; Robinson et al. [2005]; Brunelle et al. [2007]; Galbraith et al. [2008]).

The model simulates a strong direct and indirect isotope effect of denitrification. High $\delta^{15}$NO$_3^-$ produced by water column denitrification has clear regional impacts and is also responsible for overall elevated $\delta^{15}$NO$_3^-$ of the ocean relative to the N$_2$ fixation source (see below). The indirect effect of both water column and sediment denitrification is mediated by the production of N-deficient water, which creates an ecological niche for diazotrophs. This stimulates additional N$_2$ fixation when other suitable conditions for N$_2$ fixation also exist (e.g., warm (> 20°C), N-depleted water with sufficient P and Fe). This indirect effect also attenuates the horizontal circulation of high $\delta^{15}$NO$_3^-$ waters, originating from regions of water column denitrification, which causes its direct isotope effect to be regionalized to suboxic zones in the model.

Sedimentary denitrification also stimulates N$_2$ fixation, particularly over low latitude continental
shelves with high production. In the North Atlantic this effect is strong enough in the model to create a subsurface $\delta^{15}\text{NO}_3^-$ minimum, which is improves the agreement with observations. Thus, N$_2$ fixation in the ocean may be affected by the area of continental shelves (where the majority of the global sedimentary denitrification occurs), which in turn is a function of global sea level [Christensen et al., 1987; Altabet and Curry, 1989]. This suggests that glacial/interglacial changes in N$_2$ fixation in the subtropical North Atlantic may have partly been driven by sea level change and its impact on sedimentary denitrification (see also Ren et al., [2009]).

Key features of the model have been identified that are in need of further development. The coarse resolution physical circulation model does not fully resolve the dynamics of coastal upwelling regimes, which in part drive the flux of organic matter towards the sea floor sediments and its remineralization in the water column, as well as indirectly influences ventilation of suboxic zones. This is critical in the simulation of water column denitrification and sedimentary denitrification, which are important processes with respect to the global N isotope balance. Future model versions will include additional vertical levels to better resolve continental shelves. The model neglects dynamic elemental stoichiometry such as high N:P ratios of diazotrophs and the more efficient recycling of DOP relative to DON in microbial loops, which can help relieve diazotrophs of their P limitation and allow them to fix additional N$_2$ into the oceanic fixed-N pool in oligotrophic waters. The ecosystem model also suffers from the exclusion of Fe as a prognostic tracer preventing it from being able to simulate differences in ecosystems limited by macronutrients (NO$_3^-$, PO$_4^{3-}$) versus micronutrients (Fe).

Future applications of this model will include simulations of past climates, and direct comparison with $\delta^{15}$N sediment records will be used to test the model results. This approach may be a useful to quantify past interactions between the marine N cycle and its isotopes, as well as their impact on
climate and may provide new insights into important physical and biogeochemical changes throughout Earth’s history.

Appendix A. Nitrogen Isotope Model

The open system fractionation equation is used for fractionation during algal NO$_3^-$ assimilation [Altabet and Francois, 2001]:

$$
\delta^{15}P_{O} = \delta^{15}NO_3^- - \varepsilon_{ASSIM}(1 - u_{NO3}),
$$

(A1)

where $\delta^{15}P_{O}$ is the $\delta^{15}$N of phytoplankton biomass assimilated during one time step, $\Delta t$, and $u_{NO3}$ is the fraction of NO$_3^-$ available that is converted into biomass ($u_{ASSIM} = J_{OP} x \Delta t / NO_3$). When algae assimilate all available NO$_3^-$ into their biomass (i.e. $u_{ASSIM} = 1$) they will incorporate the same $\delta^{15}$N value as that of the source material. Many studies have estimated the fractionation factor in both laboratory and ocean environments. A wide variety of values have been reported in culture settings ranging from 0.7‰ to 23‰ [Wada and Hattori, 1978; Montoya and McCarthy, 1995; Waser et al., 1998; Neeboda et al., 2003; Granger et al., 2004]. A more confined range has been observed in field estimates from 4‰ to 15‰ [Wada, 1980; Altabet et al., 1991; Altabet and Fracois, 1994; Wu et al., 1997; Altabet et al., 1999; Sigman et al., 1999; Altabet and Francois, 2001; Karsh et al., 2003; DiFiore et al., 2006; Needoba et al., 2006]. In our model we choose a constant value of 5‰ which is near the majority of estimates, although it is important to bear in mind the uncertainty in the parameter choice and the possibility that it varies in space and time.

Nitrate in suboxic waters have been observed to have much higher $\delta^{15}$N values due to fractionation during denitrification. Observations from present day suboxic zones in the Eastern Tropical North Pacific (ETNP) and the Arabian Sea (AS) have reported fractionation factors ranging from 22-30‰.
[Cline and Kaplan, 1975; Liu and Kaplan, 1989; Brandes et al., 1998; Altabet et al., 1999; Voss et al., 2001; Sigman et al., 2003]; we adopt a value of 25‰ in the model. Note that because these estimates were derived from field studies in which the isotope effect was estimated from the total nitrogen loss, they implicitly include the effect of anammox [Galbraith et al., 2008]. Fractionation during denitrification is also simulated using the open system fractionation equation

\[ \delta^{15}NO_3^{\text{OX}} = \delta^{15}NO_3^- - \varepsilon_{\text{WCD}}(1 - u_{NO_3}), \]  

where $NO_3^{\text{OX}}$ is the oxygen-equivalent reduction of nitrate converted into $N_2$ gas during denitrification.

The term $u_{\text{DENI}}$ is the fraction of available $NO_3$ which is reduced into $N_2$ gas ($u_{NO_3} = \mu_D x 0.8 x R_{ON} x r_{NO_3}^{\text{sox}} x L_{NO_3} x \Delta t / NO_3$).

Excretion is the process responsible for the step-wise enrichment of $\delta^{15}N$ along the trophic chain in our model and is simulated using the instantaneous fractionation equation:

\[ \delta^{15}NO_3^- = \delta^{15}Z - \varepsilon_{\text{EXCR}}. \]  

The instantaneous fractionation equation is used because excretion will always be a small fraction of the total zooplankton biomass and has been measured to be depleted by ~6‰ relative to its body [Montoya, 2008], which is the source of the excreted nitrogen. This leads to the average enrichment of ~3.4 per trophic level [Minagawa and Wada, 1984].

Implementing these fractionation equations into the marine ecosystem model requires us to consider the exchanges of $^{14}N$ and $^{15}N$ between the various N pools separately. Total nitrogen abundance now
has the form

\[ N = {^{14}N} + {^{15}N} \]  \hspace{1cm} (A4)

for each variable in the isotope model. A fractionation coefficient is calculated for each process so the same equations for total N can be applied to \(^{15}\)N [Giraud et al., 2000]. For example, consider fractionation during algal NO\(_3^-\) assimilation. The isotopic ratio of new nitrogen biomass (\(P_O\)) is found using equations (1) and (2):

\[ ^{15}P_O = \beta_{\text{ASSIM}}^{14}P_O \]  \hspace{1cm} (A5)

where

\[ \beta_{\text{ASSIM}} = \frac{{^{15}NO_3}}{{^{14}NO_3}} - \frac{\varepsilon_{\text{ASSIM}} (1 - u_{NO_3}) R_{\text{std}}}{1000}. \]  \hspace{1cm} (A6)

Applying equations (C4) and (C5) gives the amount of new \(^{15}P_O\) relative to the amount of total new nitrogen biomass, which is given by the primary production \((J_O P_O)\), calculated by the marine ecosystem model.

\[ ^{15}P_O = \frac{\beta_{\text{ASSIM}}}{1 + \beta_{\text{ASSIM}}} J_O P_O \]  \hspace{1cm} (A7)

Analogous derivations can be done for all fractionation coefficients. The time-dependent set of equations for \(^{15}\)N which are embedded into the marine ecosystem model are as follows:
\[ \frac{\partial^{15} NO_3}{\partial t} = \left( R_{D} \mu_{D} D + \frac{\beta_{\text{EXCR}} \gamma_{2} Z + R_{P} \mu_{P} P_{O} - \beta_{\text{ASSIM}}}{1 + \beta_{\text{EXCR}}} J_{O} P_{O} - \beta_{\text{ASSIM}} J_{O} P_{O} - \beta_{\text{ASSIM}} u_{N} J_{O} D_{D} P_{O} \right) \times \left[ 1 - \frac{\beta_{\text{WCD}}}{1 + \beta_{\text{WCD}}} 0.8 R_{O,N}^{NO_3} L_{NO_3} \right] \] 

(A8)

\[ \frac{\partial^{15} P_{O}}{\partial t} = \frac{\beta_{\text{ASSIM}}}{1 + \beta_{\text{ASSIM}}} J_{O} P_{O} - R_{P} \mu_{P} P_{O} - R_{P} G(P_{O}) Z - R_{P} \mu_{P} P_{O}^{2} \] 

(A9)

\[ \frac{\partial^{15} P_{D}}{\partial t} = \left( \frac{\beta_{\text{ASSIM}}}{1 + \beta_{\text{ASSIM}}} u_{N} + \frac{\beta_{\text{NFX}}}{1 + \beta_{\text{NFX}}} (1-u_{N}) \right) J_{D} P_{D} + -R_{P} G(P_{D}) Z - R_{P} \mu_{P} P_{D} \] 

(A10)

\[ \frac{\partial^{15} Z}{\partial t} = \gamma_{1} \left[ R_{P} G(P_{O}) + R_{P} G(P_{D}) \right] Z - \frac{\beta_{\text{EXCR}}}{1 + \beta_{\text{EXCR}}} \gamma_{2} Z - R_{P} \mu_{P} Z^{2} \] 

(A11)

\[ \frac{\partial^{15} D}{\partial t} = 1 - \gamma_{1} \left[ R_{P} G P_{O} + R_{P} G P_{D} \right] Z + R_{P} \mu_{P} P_{D} + R_{P} \mu_{P} P_{O}^{2} + R_{P} \mu_{P} Z^{2} - R_{D} \mu_{P} D - R_{D} \mu_{P} D \] 

(A12)

\[ \frac{\partial^{15} D}{\partial t} = \]  

where \( R_{N=P,O,D,Z,D} = ^{15}N / (^{14}N + ^{15}N) \) is the ratio of heavy over total nitrogen. The complete parameter description is provided in Supplementary Material 2. Here it suffices to note that the equations for total nitrogen \(^{14}N + ^{15}N\) are identical to the ones of \(^{15}N\) except that \( R_{N} = \beta_{X}/(1+\beta_{X}) = 1 \) in the total nitrogen equations.

The model was carefully tested with zero fractionation in order to quantify and minimize numerical errors, which can occur for example due to slightly negative values of biological tracers caused by
inaccuracies of the advection scheme. The biological code was adjusted to avoid negative concentrations as much as possible. Initially numerical errors in $\delta^{15}$N ranged from $\pm 1$‰ in grid points at the sea floor to $\pm 0.1$‰ in the upper ocean. Setting $R_{std} = 1$ instead of $R_{std} = 0.0036765$, the actual atmospheric $N_2$ isotope ratio, reduces the numerical errors by over an order of magnitude. $R_{std}$ is set to the value 1 so both isotope variables will be on the same order of magnitude. This prevents $^{15}$N from becoming very close to zero as often, where inaccuracies of the advection scheme can cause it to be negative. This modification amounts to a scaling of $^{15}$N and $^{14}$N by a constant factor which does not affect the $\delta^{15}$N dynamics. The remaining numerical errors of $\pm 0.1$‰ in the deep ocean and $\pm 0.01$‰ in the upper ocean are two orders of magnitude smaller than the observed variability. The model is integrated for over 7,000 years as it approaches equilibrium.
References Cited


Altabet, M. A., D. W. Murray and W. L. Prell (1999), Climate-linked variations in Arabian Sea denitrification over the last 1 m.y.: Implications for the marine N cycle, *Paleoceanogr., 14,* 732-743


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Gent, P. R., and J. C. McWilliams (1990), Isopycnal mixing in ocean circulation models, *J. Phys. Oceanogr.*, 20, 150-155


Ren, H., D. M. Sigman, A. N. Meckler, B. Plessen, R. S. Robinson, Y. Rosenthal, G. H. Haug (2009), Foraminiferal Isotope Evidence of Reduced Nitrogen Fixation in the Ice Age Atlantic Ocean, Science, 323, 244-248


<table>
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<th>Field Estimates (%) $^a$</th>
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<td>4 – 15</td>
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<td>22 – 30</td>
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$^a$ see Appendix A for references

**Table 1:** Nitrogen Isotope Model Enrichment Factors
### Table 2: Global measures of $\delta^{15}$NO$_3^-$ model performance: correlation coefficient ($r$) and root mean squared ($RMS$) error normalized by the standard deviation of the observations.

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<tr>
<td>Sedimentary Denitrification</td>
<td>0.65</td>
<td>0.90</td>
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Figure 1. Schematic of the marine ecosystem model with the nitrogen isotope model parameters in color.
Figure 2. Estimated rate of annual (A) aeolian dissolved Fe deposition [Fan et al., 2006] which limits (B) N$_2$ fixation when below ~10 μmol m$^{-2}$ yr$^{-1}$ (see text). Annual vertically integrated rates of (B) N$_2$ Fixation, (C) water column denitrification, and (C) sedimentary denitrification. Contour intervals are 210 and 1000 mmol N m$^{-2}$ yr$^{-1}$.
Figure 3. Top Panel: Surface $\delta^{15}$NO$_3$ and $\delta^{15}$N of sinking detritus in the model. Bottom Panel: Isotope effect experiments where one isotope effect is neglected per simulation and its difference with CNTRL is shown to illustrate its individual effect on the CNTRL simulation.
Figure 4. Comparison of annual $\delta^{15}\text{NO}_3$ (‰) averaged between 200 m - 300 m of available observations (OBS) and CNTRL. Due to the incomplete temporal coverage, seasonal biases in the annually averaged data exist depending on the region.
Figure 5. Comparison of the Indian-Pacific sector of the Southern Ocean with the $\delta^{15}$NO$_3$ database and CTRL. (a) $\delta^{15}$NO$_3$ vs. NO$_3$; (b) horizontally averaged (over available data) depth $\delta^{15}$NO$_3$ profiles; (c) surface $\delta^{15}$NO$_3$ and with a 1$\mu$M NO$_3^-$ contour line.
Figure 6. Comparison of the ETNP with the $\delta^{15}$NO$_3$ database and CTRL. (a) $\delta^{15}$NO$_3$ vs. N' = NO$_3^-$−16PO$_4^{3-}$; (b) horizontally averaged (within 10 $\mu$M O$_2$ contour) depth $\delta^{15}$NO$_3$ profiles including the experiment where the isotope effect of N$_2$ Fixation is neglected (No NFIX); (c) subsurface $\delta^{15}$NO$_3$ with a 10 $\mu$M O$_2$ contour line.
Figure 7. Comparison of the North Atlantic with the $\delta^{15}$NO$_3$ database and CNTRL. (a) horizontally averaged (over available data) depth $\delta^{15}$NO$_3$ profiles including the experiment where sedimentary denitrification is neglected (No SD); (b) subsurface $\delta^{15}$NO$_3$. 